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1 Investigating the phenology of seaward migration of juvenile
2 brown trout (*Salmo trutta*) in two European populations
3

4 Final accepted version, ~~published in~~ *Hydrobiologia* Feb 2016

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13 Abstract

14 Recent evidence supports the existence of a downstream autumn migratory phenotype in juvenile
15 anadromous brown trout (*Salmo trutta*), however the precise timing, extent and ecological
16 significance of such behaviour remains ambiguous. We investigated the phenology of downstream
17 migration of wild juvenile trout using passive integrated transponder (PIT) telemetry over an eight
18 month period in two European rivers; the River Deerness, north-east England, and the River
19 Vilestrup, Denmark. The incidence of autumn-winter seaward migration was greater in the Deerness
20 than the Vilestrup, with at least 46 % of migrating juveniles detected prior to spring smoltification in
21 the Deerness. Timing of migration was strongly regulated by factors associated with river discharge
22 in both systems. While autumn and spring downstream migrants did not differ in size at the time of
23 tagging in either system, evidence that spring migrants were of better condition, travelled faster
24 (autumn: 11.0 km day⁻¹; spring: 24.3 km day⁻¹) and were more likely to leave the Deerness suggests
25 autumn and spring migrant conspecifics respond to different behavioural motivations. Further
26 investigation into the sex of autumn migrant juveniles, as well as the temporal and geographical
27 variability in the incidence and fitness consequences of autumn emigration by juvenile trout would
28 be beneficial.

29 Keywords: Autumn migration, smolt, ~~Salmo trutta~~, life history, phenotype, phenology, telemetry

30 Introduction

31 Individuals of migratory species often exhibit wide variation in their spatial behaviour, varying
32 from local residency to large-scale migration (Nathan et al. 2008; Chapman et al. 2011;~~Dingle,~~
33 ~~2014~~), including for fishes (Lucas & Baras, 2001). The literature concerning such variation within the
34 Salmonidae is voluminous (~~reviewed~~ in Dodson et al. 2013). The brown trout, *Salmo trutta* L.,
35 exemplifies such phenotypic plasticity with its spatial and temporal continuum of alternative
36 migratory tactics, from locally-resident to potamodromous and anadromous forms (Jonsson &
37 Jonsson, 2011; Boel et al. 2014). Partial and differential migration, at the intrapopulation level, have
38 been illustrated in numerous *S. trutta* populations, notably around Vangsvatnet Lake, Norway
39 (Jonsson, 1985), in the Oir River, France (Cucherousset et al. 2005) and with regard to the extent of
40 marine migration (~~del~~ Villar-Guerra et al. 2013; Aarestrup et al. 2014).

41 Despite thorough accounts of behaviour, elements of juvenile salmonid space use and
42 phenology of downstream migration remain poorly explained. It is traditionally considered that
43 juvenile populations of anadromous brown trout and closely related Atlantic salmon, *S. salar*, remain
44 in their native stream from one to several years before descending the river course as smolts in
45 spring (Klemetsen et al. 2003). Therefore, when considering anadromous salmonid population
46 dynamics, the freshwater output of a river typically refers to the production of spring migrants
47 (Ibbotson et al. 2013). Estimates of juvenile salmonid population density are usually carried out in
48 late spring when site fidelity is regarded as strong (Cunjak 1992) and one is typically unable to
49 distinguish mortality from emigration when examining population losses over autumn and winter.

50 Few published tracking or trapping studies have targeted juvenile trout outside spring (but see
51 Jonsson & Jonsson, 2002, 2009; Marine Institute, 2014; Holmes et al. 2014; Taal et al. 2014). For
52 example, in widely cited studies on trout migration, Elliott (1986, 1994) did not investigate evidence
53 that small-scale emigration of Age 1+ trout parr from Black Brows Beck, NW England, appeared to
54 occur year round and represented a high proportion, relative to spring smolt numbers.

Comment [EW1]: I'm not sure whether this sentence is a little redundant.

Comment [M2]: Emily, leave this in. It is a real issue and problem – that was discussed specifically at a meeting I was recently. So keep that sentence in please.

55 Downstream movements of *S. salar* juveniles are well known in autumn (Youngson et al. 1983;
56 Cunjak et al. 1989; Pinder et al. 2007; McGinnity et al. 2007; Jensen et al. 2012; Jonsson & Jonsson,
57 2014; Taal et al. 2014), although the proportions of populations undertaking those movements, and
58 their significance, ~~are~~ still not fully understood. By contrast, autumn descents of immature brown
59 trout have been less widely recorded, but information is available in Irish (Marine Institute, 2014),
60 Norwegian (Jonsson & Jonsson, 2002, 2009), Baltic (Taal et al. 2014) and New Zealand (Holmes et al.
61 2014) populations. Based upon population-specific interpretations of phenotype, non-moribund,
62 juvenile salmonid autumn migrants are regarded as pre-smolts, destined to enter the sea (McGinnity
63 et al. 2007), or as comprising potamodromous population components that may be common in
64 larger river systems (Cucherousset et al. 2005). Survival may also differ by population, for example
65 the weakly brackish environment encountered by autumn migrants in the Baltic Sea does not
66 require marine-level osmoregulatory competency for survival (Taal et al. 2014). ~~Alternatively, but~~
67 ice, low river discharge and the absence of an estuary in which to reside are thought to cause poor
68 survival of *S. salar* autumn migrants leaving the River Halselva, Norway and a lack of sympatric *S.*
69 *trutta* conspecifics (Jensen et al. 2012). Autumn migrant *S. salar* do not appear suitably adapted to
70 seawater (Riley et al. 2008), and sea survival rates of first-time, autumn-emigrant *S. trutta* in Norway
71 are significantly lower than those in spring (Jonsson & Jonsson, 2009), however the viability of these
72 phenotypes has been confirmed with returning *Salmo* adults (Jonsson & Jonsson, 2009; Riley et al.
73 2009).

74 Despite enhanced awareness of an autumn downstream migratory phenotype in juvenile
75 salmonids over recent years, our understanding of its ecological significance remains limited and
76 somewhat contested. Recorded incidence of autumn migration has been associated with proximity
77 to the marine environment (Ibbotson et al. 2013), elevated stream discharge (Youngson et al. 1983)
78 and poor overwintering habitat (Riddell & Leggett, 1981), as well as the reproductive motivations of
79 precociously mature male parr (Buck & Youngson, 1982; McGinnity et al. 2007; Jensen et al. 2012).
80 Holmes et al. (2014) suggested that early emigration of larger trout parr from the Rainy River, New

81 Zealand reflected limited overwintering habitat availability and/or constrained feeding opportunities
82 for individuals with high growth rates. Determining the value of alternative wintering habitats, in
83 terms of trout survival, smolt output, performance in the sea and ultimately their fecundity , is
84 important for river and fisheries managers in directing conservation, regulation and habitat
85 management (Ibbotson et al. 2013), such as when considering the importance of year-round
86 downstream fish passage.

Comment [EW3]: Maybe I'm being too ruthless by cutting this...

Comment [M4]: I agree this sentence could be cut

87 In this study we examined and compared the phenology of downstream migration in two
88 juvenile trout populations in north-east England and Denmark, using Passive Integrated Transponder
89 (PIT) telemetry. We investigated the influence of environmental, subject-specific and tag-site specific
90 variables on the probability of passage past fixed monitoring sites over an eight month period.

91 **Materials and Methods**

92 *Study areas*

93 The River Deerness (mean annual discharge $\sim 0.5 \text{ m}^3 \text{ s}^{-1}$ in its lower reaches) is a small tributary
94 (width in study reaches, 2-10 m) of the River Browney in NE England ($54^\circ 44' \text{ N}$, $1^\circ 48' \text{ W}$) and flows
95 into the North Sea via the River Wear (Fig. 1). The Wear has large stocks of Atlantic salmon and sea
96 trout with annual resistivity (partial) counts of upstream adult migrants at Durham, 29 km from the
97 sea, averaging 15,593 fish per year between 2007 and 2014 (Environment Agency, 2015⁴). The
98 Deerness drains a catchment of 52.4 km^2 , and is 16.3 km long, consisting largely of grassland to the
99 west, and transitioning to woodland and lower-lying arable areas in the east. The brown trout
100 population consists of multiple cohorts, with a mixture of resident and migratory adults, including
101 anadromous trout (E. Winter, J. Tummers unpublished data).

102 The River Villestrup ($56^\circ 46' \text{ N}$, $9^\circ 55' \text{ E}$) is the primary freshwater source for the strongly
103 brackish Mariager Fjord, ultimately exchanging with the Kattegat Sea on the east coast of Jutland,
104 Denmark (Fig. 1). The river has a mean annual discharge of $1.1 \text{ m}^3 \text{ s}^{-1}$. The Villestrup is approximately

20 km long, has a typical width in the studied reach of 4-10 m and drains a catchment of 126 km². The average density of wild 0+trout is estimated at 125 per 100 m² (HELCOM, 2011). No stocking occurs. The inner fjord has salinities of 12-17 PSU in the upper 10 m of the water column used by trout, while deeper areas are more saline but often hypoxic. The shallow outer fjord has salinities of 20-25 PSU. The Villestrup joins near the junction between the inner and outer fjord areas, on the north shore (Fig. 1).

PIT tagging, recapture and telemetry

Trout in the Deerness system were captured for PIT-tagging using electric-fishing equipment at six sites dispersed over *ca.* 15 km of stream length (Fig. 1). Tagging occurred between 9 July and 12 September 2014 (*n* = 643), with a small number (*n* = 23) also tagged on 6 November 2014 (mean FL \pm SD of all trout = 151 mm \pm 23). In the Villestrup, trout were tagged on 26 September 2014 (*n* = 490; mean FL \pm SD = 147 mm \pm 27), using electric-fishing in a single reach *ca.* 8.5 km upstream of the river outlet (Fig. 1). Parr \geq 120 mm (Larsen et al. 2013) and \leq 250 mm from each study area were anaesthetised (Deerness: buffered MS-222, 100 mg L⁻¹; Villestrup: Benzocaine, 25 mg L⁻¹), weighed (to 0.1 g), measured (fork length, FL to 1 mm) and surgically implanted with a PIT-tag (Texas Instruments; model RI-TRP-RRHP, HDX, 134.2 kHz, length 23.1 mm, diameter 3.85 mm, weight 0.6 g in air). Tags and instruments were disinfected with 90% ethanol and air dried before use. Procedures were carried out by an experienced fish surgeon and following local animal welfare regulations. Following recovery, all individuals were returned to their site of capture. Recapture methodology used to investigate summer dispersal on the Deerness is presented in Online Resource 1.

Three pairs of stream-width swim-through half-duplex (HDX) PIT antennae were installed on a 5 km stretch of the lower Deerness (Fig. 1; Bolland et al. 2009, 2010), operational from 24 September 2014 at stations M2 and M3 and 13 October at station M1 (Fig. 1), until 31 May 2015. Due to occasional battery failure and a damaging high flow event in November causing severe loss of efficiency, the stations M1, M2 and M3 were operational 95.6, 98.1 and 98.1 % of the time,

respectively. On the Villestrup, a single pair of mains-powered antennae were placed 300 m upstream of the river outlet (Fig. 1), functional 88.7 % of the time from 26 September 2014 to 31 May 2015. Tags were detected by time-synchronised Master and Slave HDX readers (Texas Instruments SX2000; in-house build), interrogating the pairs of antennae eight times per second (Castro-Santos et al. 1996). Detection ranges between 20 and 80 cm were achieved and correct function on the Deerness was confirmed by passing a test tag through each antenna before and after each battery change (every 4 ± 2 days), and by more detailed range testing periodically. Each Villestrup antenna had a timed auto-emitter check tag (Oregon RFID). Additionally on the Villestrup, a Wolf-type trap (Wolf, 1951) situated directly downstream of the PIT antennae captured migrating smolts from 18 March 2015 until the end of the study. For the purpose of this study, and by reference to standard terminology applied elsewhere (e.g. Ibbotson et al. 2013), all fish detected prior to 1 February were labelled autumn migrants, while those detected from 1 February to 31 May were labelled spring smolts.

Theoretical antenna efficiencies of 99, 100 and 100% were achieved for stations M1-3, respectively, by routinely passing a test tag through the system. In practice, several factors influence a tag's probability of detection, including environmental conditions, tag velocity, tag orientation and the presence of other tags (Zydlewski et al. 2006; Burnett et al. 2013). A practical estimate of efficiency is, hence, given by the ratio of fish detected at a site that are known to have passed through (Zydlewski et al. 2006), and was estimated at 98.4% for M2. The efficiencies of M1 and M3 could not be estimated using this method, due to the absence of detection equipment downstream and the time lag between tagging events and the onset of monitoring, meaning the location of individuals prior to detection was uncertain. Using Zydlewski et al.'s (2006) method, the efficiency of the Villestrup station was estimated as 86.5% in spring, by identifying individuals caught in the downstream trap that were not detected by the PIT antennae.

Population density estimations and environmental monitoring

Quantitative estimates of trout densities ($n \cdot 100 \text{ m}^{-2}$) were made at each of the Deerness tagging sites, using a multiple-pass depletion method (Carle & Strub, 1978) in July 2014 (average triple-pass catchability 97.2% for \geq Age 1) and subsequently in March 2015 (average triple-pass catchability 98.4% for \geq Age 1). Developmental state of recaptured individuals (parr, smolt [including part-smolt] or adult) was also recorded, based on phenotypic characteristics (e.g. parr marks, body colour, body shape; Tanguy et al. 1994), in order to predict the seaward movement of certain individuals.

One logger (HOBO®; model U20-001-01; Onset Computer Corporation), situated at M3 on the Deerness, recorded temperature (accuracy $\pm 0.4 \text{ }^{\circ}\text{C}$) and water pressure (accuracy $\pm 0.6 \text{ kPa}$, converted to river level) at 15-min intervals throughout the study. On the Villestrup, temperature was measured at the upper antenna (Tinytag plus 2; model TGP-4017, www.geminidataloggers.com) and water level records were obtained from a fixed gauging station ca. 1 km upstream of the river outlet (Fig. 1).

Statistical analyses

The influence of environmental factors on the timing of downstream movement was analysed using generalised linear models (GLMs), comparing daily detection frequency with fluctuations in average daily water temperature ($^{\circ}\text{C}$), photoperiod, average daily water level (m) and the change in average daily water level (m). Initial Poisson GLMs revealed non-linear residual patterns and overdispersion, which was corrected for using the negative binomial distribution with a log-link function (Richards, 2008). All combinations of explanatory variables were trialled, but never utilising collinear factors (e.g. temperature and photoperiod). Following this, we explored the effects of subject-specific variables on the probability of tagged trout being detected as autumn or spring migrants. Binomial GLMs with a cloglog link function, due to asymmetry in the numbers of migrants and non-migrants, were constructed using combinations of the independent factors fish length (mm), mass (g) and Fulton's condition factor, but never with collinear variables (e.g. length and

mass). The additional variables tag-site density ($n \cdot 100m^{-2}$, summer 2014), tag-site distance upstream (km), and date of tagging were also analysed for subjects on the Deerness. Twelve Deerness fish, one a spring migrant, one an upstream migrant and ten undetected, and two Villestrup fish, one a spring migrant and one undetected, were omitted from this analysis due to a lack of mass, and hence condition, records. All analyses were conducted in R 3.1.1 (R Core Team, 2014) with use of the MASS package (Venables & Ripley, 2002). Model selection followed the minimisation of Akaike's information criterion (AIC) values, which represents the best compromise between lack of precision (too many parameters) and bias (too few parameters). Models within $\Delta 6$ AIC were retained, provided they were not increasingly complex versions of more efficient nested counterparts (outlined by Richards, 2008).

For further examination, we used independent-sample *t*-tests to compare the average length, mass, condition factor and net ground speed ($km \ day^{-1}$) of autumn and spring migrating individuals. Chi-squared tests with Yates' continuity correction were used to determine if the proportion of Deerness fish caught, inspected and categorised as smolts in March and subsequently detected downstream differed by tag site, and if the proportion of Deerness downstream migrants reaching M1 differed between autumn and spring.

Results

Site fidelity was strong during summer 2014, with 83% of recaptured Deerness trout parr ($n = 330$) travelling no further than 60 m and $< 0.01\%$ travelling over 200 m, from a previous known site of release (Online Resource 2). From autumn onwards, 140 (21.0%) of the 666 Deerness trout were PIT detected downstream of the site at which they were tagged and released, comprising 83 autumn migrants, 52 spring migrants and five individuals with activity spanning both periods (from here on labelled autumn migrants). Trout detected at M1 were assumed to have left the Deerness system, comprising 89 individuals in total (13.4%), 41 in autumn and 48 in spring. Migratory behaviour was observed throughout the period of study, with peaks of activity in October, November, March and

May (Fig. 2). Eleven upstream migrants from T1 to M3 were recorded, all occurring in autumn and none of which were detected leaving the stream. In March 2015, parr densities had decreased at all but one of the tagging sites (Table 1), by an overall average of 49%; a total of 50 tagged trout were recaptured in March across all release sites, of which 17 subsequently migrated downstream.

In the Villestrup, 195 of 490 tagged trout (39.8 %) were detected at the monitoring site and/or caught in the trap, comprising 49 autumn migrants, 136 spring migrants, and ten individuals with activity spanning both periods (from here on labelled autumn migrants). The Villestrup trout did not display a distinct autumnal peak of activity, however low levels of migratory behaviour were sustained throughout autumn and winter months. Activity of spring smolts peaked in late March, with continued high levels of movement throughout April and early May (Fig. 2).

Environmental regulators of downstream migratory behaviour

Mean daily water temperature and water level were retained in the best model for predicting autumn migrant activity in the Deerness (Table 2), both sharing significant positive relationships with daily detection frequency. No other combinations of variables were retained under the selection criteria, reflecting their poor explanatory power. In contrast, the change in mean daily water level was the best predictor of spring migrant activity in the Deerness, with a significant positive relationship. Two further models were retained utilising the variables water level and photoperiod, however greater ΔAIC values signified weaker explanatory power (Table 2). The best model predicting autumn migrant activity in the Villestrup retained both water level and temperature, but, in contrast to the Deerness, with a significant negative effect of temperature (Table 2). Under the selection criteria, five models were retained for predicting spring migrant activity in the Villestrup, with the best predictors being change in mean daily water level and water temperature in positive trends (Table 2). The removal of temperature produced a model with a ΔAIC of 0.3, indicating the explanatory power of temperature is low and water level on its own is a powerful predictor of daily smolt counts.

Phenotypic determinants of downstream migratory tendency

The best model describing autumn migrant probability in the Deerness retained fish mass and tag site distance upstream as predictive parameters, both with significant negative trends (Table 3; Fig. 3). Three alternative models utilising combinations of length, condition, distance and date can be found in Table 3. The best model for predicting spring migration retained subject mass, condition factor and tag site distance upstream. Mass and tag site distance shared significant negative relationships with probability of spring detection, while condition factor shared a positive relationship (Table 3; Fig. 3). Notably, tag site distance was retained in all models for the Deerness, suggesting it is a strong predictor of migratory tendency for all downstream migrating juveniles, but particularly for spring smolts, given the higher coefficient estimates. This is supported by significantly more smolts (captured, inspected and classified in March 2015) than expected being detected originating from T1, the most downstream tag site, relative to sites further upstream (Chi-square: $\chi^2_1 = 6.18$, $p = 0.013$). Autumn and spring migratory tendency in the Villestrup were best predicted by fish mass and condition, both sharing negative relationships in each scenario (Table 3; Fig. 3). For the autumn migrants, mass was a particularly strong predictor on its own, given the removal of condition to produce a model with a ΔAIC of only 0.7.

Comparisons of migratory phenotypes

There was no difference in length ($t_{138} = 0.22$, $p = 0.82$) or mass ($t_{137} = 0.39$, $p = 0.70$) at the time of tagging between autumn and spring migrants in the Deerness; however there was some evidence to suggest condition factor was greater in spring migrants ($t_{137} = 2.00$, $p = 0.048$; Fig. 3). Mean length and mass of upstream migrants was also significantly greater than that of all downstream migrants in the Deerness (Length: $t_{149} = 3.55$, $p < 0.001$; Mass: $t_{147} = 4.62$, $p < 0.001$; Fig. 3), although we found no difference in condition factor ($t_{147} = 0.05$, $p = 0.96$). There was no difference in length ($t_{193} = 0.12$, $p = 0.90$), mass ($t_{192} = 0.26$, $p = 0.79$) or condition factor ($t_{192} = 0.32$, $p = 0.75$; Fig. 3) between autumn and spring migrants in the Villestrup. Individual net ground speed of downstream migrants in the

Deerness varied dramatically from less than 1 to 88 km day⁻¹ throughout the study period, but on average, spring migrants travelled significantly faster (24.3 km day⁻¹) than autumn migrants (11.0 km day⁻¹) ($t_{127} = 3.82$, $p < 0.001$; Fig. 4). Additionally, downstream migrants were classified as stream 'emigrants' following a final detection at the most downstream monitoring site, *ca.* 700 m from the Deerness' confluence with the river Browney. A significantly greater proportion of spring migrants (84.6%) than autumn migrants (49.4%) became stream emigrants during the period in which they were first detected (Chi-square: $\chi^2_1 = 15.5$, $p < 0.001$). Five autumn non-emigrants were subsequently detected in the spring, four of which were then detected moving past M1.

Discussion

Extensive autumn downstream migrations were evident in juvenile brown trout (*Salmo trutta*) for both the Villestrup (Jutland, Denmark) and the Deerness (north-east England). Over 1.5 times the number of spring smolts were observed active between October and January in the Deerness. Moreover those leaving the system during autumn-winter represented 46% of all stream-emigrant juveniles in the study period. This contrasts the strong site fidelity observed during summer months and provides quantitative evidence of the protracted overwinter nature of the downstream migratory behaviour, broadly similar to that of the Marine Institute (2014), which states numbers of autumn migrating juveniles in the Burrishoole catchment, Ireland, have fluctuated between 18% and 57% of the total annual juvenile downstream migrant count, since 1982. Both the Burrishoole and Deerness catchments are characterised by mild, oceanic climates and frequent flow elevations following rainfall, possibly accounting for the prolonged 'autumn' downstream migration. Nevertheless, Deerness migrants cannot fully account for the marked reduction in parr densities at sites T2-T5 by March 2015, which must also reflect either local movement outside tagging sites, low overwinter survival, or a combination of both.

Probability of migration was higher in the Villestrup, but incidence of autumn migration was lower (25% of the total juvenile stream-emigrant count), yet remains higher than that reported by

280 Jonsson and Jonsson (2009) in Norway. Migrants in the Villestrup are assumed to enter the brackish
281 Mariager Fjord soon after passage through the PIT monitoring station, however autumn migrants
282 may have low gill Na^+K^+ -ATPase activity, as observed for autumn-emigrating *S. salar* juveniles (Riley
283 et al. 2008). This could cause osmoregulatory stress, although salmonid parr have been known to
284 reside in estuarine environments (Cunjak et al. 1989; Pinder et al. 2007). Conversely, individuals in
285 the Deerness have much further to travel before reaching a saline environment, (meaning their
286 migration strategy, i.e. anadromy or potamodromy, is unknown), and may explain why incidence of
287 autumn migration is particularly high there. The recognition of an alternative juvenile downstream
288 migratory phenotype is growing internationally and, hence, the potential contribution of these
289 individuals to adult recruitment must be acknowledged.

290 The environmental factors regulating the phenology of downstream movement of salmonid
291 smolts have been well studied, particularly the behavioural responses to water temperature, flow
292 and light (e.g. Jonsson, 1991; Aarestrup et al. 2002; Aldvén et al. 2015). Stimuli for migration differ in
293 their importance geographically between river systems and temporally between years (Hembre et
294 al. 2001). This study suggests that water level (and hence river discharge) had the greatest influence
295 on autumn migrant movement in both the Deerness and Villestrup, in accordance with Youngson et
296 al. (1983), Jonsson and Jonsson (2002) and Holmes et al. (2014). This is not surprising, given the
297 opportunity to minimise the energetic costs of migration, while high turbidity may offer greater
298 protection from predators (Hvidsten & Hansen, 1989). ~~In contrast, the~~ change in average daily
299 water level was a better predictor of movement of spring migrants in both the Deerness and
300 Villestrup. This suggests smolts in both systems are particularly receptive to dynamic fluctuations in
301 the hydrograph, similar to Carlsen et al.'s (2004) conclusions that migrating juveniles can anticipate
302 floods. Importantly, all models retained in Table 2 utilised an environmental variable associated with
303 river discharge, suggesting it is a central migratory stimulus for both populations.

Autumn migrants in the Deerness and smolts in the Villestrup responded positively to higher temperature, analogous to results obtained by Jonsson and Ruud-Hansen (1985). Smolt activity in the Deerness was better predicted by photoperiod, which is known to regulate physiological changes associated with the parr-smolt transformation (Björnsson et al. 2011). The probability of autumn migration in the Villestrup shared a significant negative relationship with temperature (see also Jonsson & Jonsson, 2002), yet previous studies reporting increased migrant activity at cold temperatures are usually associated with ice melt (Hesthagen & Garnås, 1986; Carlsen et al. 2004). One possibility for the observed pattern is that peak flows may have coincided with low temperatures in winter, demonstrating a degree of behavioural independence with regard to seasonal variables. Also, the Villestrup is spring-fed, meaning temperatures are comparatively more stable to those of the Deerness. The relative importance of temperature, photoperiod and river discharge as migratory triggers may, however, fluctuate between years (Jensen et al. 2012; Aldvén et al. 2015), depending on precipitation and rate of temperature change. Longer-term and experimental studies in either system could reveal variability and mechanisms in the effect of environmental cues.

The probability of an individual conducting autumn or spring migration past fixed points on the Deerness decreased significantly in an upstream direction. Ibbotson et al. (2013) proposed a re-distribution of autumn migrating *S. salar* parr in a downstream direction was responsible for similar findings, rather than a targeted migration. One explanation for this may be to consider the potential disturbance caused by environmental events such as high autumn-winter flows. Territorial behaviour may be disrupted, initiating a re-establishment of dominance hierarchies and promoting the downstream displacement of subordinate individuals. While this may hold true for autumn-winter migrants, it is unlikely to be the case for spring smolts, but for which we found a reduced probability of detection from tag sites further upstream. Moreover, there appears no trend in the percentage of recaptured residents with distance upstream (Table 1), which would be expected if migration tendency decreased in an upstream direction. We, therefore, suggest a cumulative increase in

mortality probability with increasing distance upstream for both autumn and spring migrants, for example due to anthropogenic obstruction during migration, greater energetic costs or exposure to predators (Aarestrup & Koed, 2003; Bohlin et al. 2001; Gauld et al. 2013).

In early studies it was hypothesised that anadromous salmonid juveniles migrate at the first opportunity after reaching a threshold size (Elson, 1957; Fahy, 1985), yet for brown trout, smolt length can vary from <100 mm to >200 mm within the same river (Økland et al. 1993). Fast-growing individuals smolt at a younger age and smaller size than their slow-growing counterparts (Forseth et al. 1999). In the Deerness and Villestrup, autumn and spring migratory tendency were negatively affected by body mass, which was always a better predictor than body length. This may be counterintuitive, considering migration and post-migration survival is thought to be positively size-dependent (Bohlin et al. 1993), but could suggest migrating juveniles in these river systems were energetically constrained. Migration has been described as a biological response to adversity (Taylor & Taylor, 1977), and in the Deerness (autumn migrants) and Villestrup (all migrants) migratory tendency was negatively correlated with fish condition at tagging, which could result from adversity in the form of poor growth opportunities. Notably, the optimal size at migration in the Deerness and Villestrup systems could be lower than the size range of individuals sampled such that some of the tagged trout, especially males, may have been maturing.

The size of autumn and spring migrants did not differ at the time of tagging in either the Deerness or Villestrup, as found by Ibbotson et al. (2013) for *S. salar*. In contrast, some found autumn migrants to be significantly larger than spring migrants or residents of the same year-class, possibly indicating constrained habitat availability for faster-growing individuals or achievement of a high energy store status (Huntingford et al. 1992; Holmes et al. 2014). This hypothesis is not supported by our results. In the Deerness, spring migrants were of better condition than autumn migrants at the time of tagging and autumn and spring migratory tendencies shared opposed relationships with individual body condition. This may have been a result of competition, such that

Comment [M5]: No, I think leave all of this and the references in – I think this is a significant (popular) area of debate and it is relevant to include

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subordinate individuals with lower body condition were displaced by their dominant counterparts, either preferentially leaving a low-growth potential environment or forced out of refugia with an overwinter reduction in stream carrying capacity (see Keeley, 2001). Migration in brown trout can be regulated by food availability (Wysujack et al. 2008), therefore autumn migrants could result from a competition-induced lack of resources for subordinates. On the contrary, we found no evidence to suggest the density of trout Age 1+ and older at each Deerness site could predict the proportion of autumn migrants. However, caution is needed, since local habitat has been shown to influence the autumnal movement of Atlantic salmon parr, perhaps due to differences in the proximity of overwintering habitat (Ibbotson et al. 2013).

Autumn upstream migrants in the Deerness, characteristic of precocious parr maturation (McCormick et al. 1998), were larger than downstream migrants. Precocious parr are predominantly males and the anadromous emigrants' sex ratio is typically heavily skewed towards females (Klemetsen et al. 2003) although autumn downstream migration has been linked to the reproductive motivations of precocious parr (Buck and Youngson, 1982). ~~E, but fish~~ sex was unknown in this study, although 22.5% of tagged and untagged parr morphotypes ≥ 120 mm, sampled on the Deerness in November 2014 were spermiating males (E. Winter, unpublished data). Cheap molecular methods for the sexing of juvenile salmonids from tissue samples are now available (Quéméré et al. 2014) and will aid sex-specific interpretation of movement patterns in parr morphotype salmonids.

Questions regarding the ecological significance of autumn migration of juvenile salmonids remain. Increased plasma thyroxine levels in autumn migrants are suggestive of a physiologically mediated migration (Riley et al. 2008; Zydlewski et al. 2005), however the behavioural motivations of autumn and spring migrants may differ, since autumn-migrating parr are not physiologically adapted for seawater entry (Riley et al. 2008). Deerness spring migrants travelled at a greater net ground speed, at rates similar to those obtained by Aarestrup et al. (2002) for radio tagged trout smolts, and were more likely to become stream emigrants than autumn migrants. While antennae

malfunction during a major spate in November 2014 may be partially responsible for a lack of autumn detections at M1, these results suggest the movements of Deerness autumn individuals are not exclusively marine-targeted. In the Villestrup it seems likely that downstream-migrating parr recorded at the PIT station at the bottom of the river subsequently enter the Mariager Fjord, though they could remain in the river outlet. It would, therefore, be valuable to determine the range of salinities and temperatures to which juvenile autumn emigrants are exposed and their survival in relation to their physiological readiness for seawater transition.

In conclusion, this study provides quantitative evidence for considerable autumn and winter downstream migration of juvenile brown trout in the rivers Deerness and Villestrup, along with correlative information on factors regulating behaviour, to parallel the wealth of knowledge concerning spring smolt migration. We propose different behavioural motivations for autumn and spring migrants, based on individual condition, the proportion of emigrants and rates of migration for each group in the Deerness. We emphasise the dynamic behavioural nature of the brown trout and suggest the autumn-migratory phenotype represents an important avenue within the migration continuum concept (Cucherousset et al. 2005; Dodson et al. 2013; Boel et al. 2014), dependent on environmental and physiological factors relating to individual fitness. An in-depth, experimental evaluation of the temporal, spatial and genetic variability of the extent of autumn migration of trout, and its influence on subsequent life history traits, would be beneficial to future conservation and management plans.

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References

404 [Aarestrup, K. & A. Koed, 2003. Survival of migrating sea trout \(*salmo trutta*\) and Atlantic salmon](#)
 405 [\(*Salmo salar*\) smolts negotiating weirs in small Danish rivers. Ecology of Freshwater Fish 12: 169-176.](#)

406 Aarestrup, K., C. Nielsen & A. Koed, 2002. Net ground speed of downstream migrating radio-tagged
 407 Atlantic salmon (*Salmo salar* L.) and brown trout (*Salmo trutta* L.) smolts in relation to
 408 environmental factors. Hydrobiologia 483: 95-102.

409 Aarestrup, K., H. Baktoft, A. Koed, D. del Villar-Guerra & E. B. Thorstad, 2014. Comparison of the
 410 riverine and early marine migration behaviour and survival of wild and hatchery-reared sea trout
 411 *Salmo trutta* smolts. Marine Ecology Progress Series 496: 197-206.

412 Aldvén, D., E. Degerman & J. Höjesjö, 2015. Environmental cues and downstream migration of
 413 anadromous brown trout (*Salmo trutta*) and Atlantic salmon (*Salmo salar*) smolts. Boreal
 414 Environment Research 20: 35-44.

415 Björnsson, B. T., S. O. Stefansson & S. D. McCormick, 2011. Environmental endocrinology of salmon
 416 smoltification. General and Comparative Endocrinology 170: 290-298.

417 Boel, M., K. Aarestrup, H. Baktoft, T. Larsen, S. S. Madsen, H. Malte, C. Skov, J. C. Svendsen & A.
 418 Koed, 2014. The physiological basis of the migration continuum in brown trout (*Salmo trutta*).
 419 Physiological and Biochemical Zoology 87: 334-345.

420 Bohlin, T., C. Dellefors & U. Faremo, 1993. Optimal time and size for smolt migration in wild sea trout
 421 (*Salmo trutta*). Canadian Journal of Fisheries and Aquatic Sciences 50: 224-232.

422 Bohlin, T., J. Pettersson & E. Degerman, 2001. Population density of migratory and resident brown
 423 trout (*Salmo trutta*) in relation to altitude: evidence for a migration cost. Journal of Animal Ecology
 424 70: 112-121.

425 Bolland, J. D., I. G. Cowx & M. C. Lucas, 2009. Dispersal and survival of stocked cyprinids in a small
 426 English river: comparison with wild fish using a multi-method approach. *Journal of Fish Biology* 74:
 427 2313-2328.

428 Buck, R. J. G. & A. F. Youngson, 1982. The downstream migration of precociously mature Atlantic
 429 salmon, *Salmo salar* L. parr in autumn; its relation to the spawning migration of mature adult fish.
 430 *Journal of Fish Biology* 20: 279-288.

431 Burnett, N. J., K. M. Stamplecoskie, J. D. Thiem & S. J. Cooke, 2013. Comparison of detection
 432 efficiency among three sizes of half-duplex passive integrated transponders using manual tracking
 433 and fixed antenna arrays. *North American Journal of Fisheries Management* 33: 7-13.

434 Carle, F. L., & M. R. Strub, 1978. A new method for estimating population size from removal data.
 435 *Biometrics* 34: 621-630.

436 Carlsen, K. T., O. K. Beg, B. Finstad & T. G. Heggberget, 2004. Diel periodicity and environmental
 437 influence on the smolt migration of Arctic char, *Salvelinus alpinus*, Atlantic salmon, *Salmo salar*, and
 438 sea trout, *Salmo trutta*, in northern Norway. *Environmental Biology of Fishes* 70: 403-413.

439 Castro-Santos, T., A. Haro & S. Walk, 1996. A passive integrated transponder (PIT) tag system for
 440 monitoring fishways. *Fisheries Research* 28: 253-261.

441 Chapman, B. B., C. Bronmark, J. A. Nilsson & L. A. Hansson, 2011. The ecology and evolution of
 442 partial migration. *Oikos* 120: 1764-1775.

443 Cucherousset, J., D. Ombredane, K. Charles, F. Marchand & J.-L. Baglinière, 2005. A continuum of life
 444 history tactics in a brown trout (*Salmo trutta*) population. *Canadian Journal of Fisheries and Aquatic*
 445 *Sciences* 62: 1600-1610.

446 Cunjak, R. A., 1992. Comparative feeding, growth and movement of Atlantic salmon (*Salmo salar*)
 447 parr from riverine and estuarine environments. Ecology of Freshwater Fish 1: 26-34.

448 Cunjak, R. A., E. M. P. Chadwick & M. Shears, 1989. Downstream movements and estuarine
 449 residence by Atlantic salmon parr (*Salmo salar*). Canadian Journal of Fisheries and Aquatic Sciences
 450 46: 1466-1471.

451 ~~del~~ Villar-Guerra, D., K. Aarestrup, C. Skov & A. Koed, 2014. Marine migrations in anadromous
 452 brown trout (*Salmo trutta*). Fjord residency as a possible alternative in the continuum of migration
 453 to the open sea. Ecology of Freshwater Fish 23: 594-603.

454 ~~Dingle, H., 2014. Migration: The Biology of Life on the Move, 2nd edition. Oxford University Press.~~

455 Dodson, J. J., N. Aubin-Horth, V. Thériault & D. J. Páez, 2013. The evolutionary ecology of alternative
 456 migratory tactics in salmonid fishes. Biological Reviews 88: 602-625.

457 Elliott, J. M., 1986. Spatial distribution and behavioural movements of migratory trout *Salmo trutta*
 458 in a Lake District stream. Journal of Animal Ecology 56: 83-98.

459 Elliott, J. M., 1994. Quantitative ecology and the brown trout. Oxford University Press, Oxford.

460 Elson, P. F., 1957. The importance of size in the change from parr to smolt in Atlantic salmon.
 461 Canadian Fish Culturist 21: 1-6.

462 Environment Agency, 2015. River Wear monthly upstream fish counts.
 463 <https://www.gov.uk/government/statistics/river-wear-monthly-upstream-fish-counts>

464 Fahy, E., 1985. Cyclic fluctuations in the abundance of trout (*Salmo trutta* L.). Archiv fuer
 465 Hydrobiologie Supplement 70: 404-428.

466 Forseth T., T. F. Næsje, B. Jonsson & K. Hårsaker, 1999. Juvenile migration in brown trout: a
 467 consequence of energetic state. Journal of Animal Ecology 68: 783-793.

468 Gauld, N. R., R. Campbell & M. C. Lucas, 2013. Reduced flows impact salmonid smolt emigration in a
 469 river with low-head weirs. *Science of the Total Environment* 458-460: 435-443.

470 HELCOM, 2011. Sea Trout and Salmon Populations and Rivers in Denmark – HELCOM assessment of
 471 salmon (*Salmo salar*) and sea trout (*Salmo trutta*) populations and habitats in rivers flowing to the
 472 Baltic Sea. Baltic Sea Environment Proceedings No. 126B.

473 Hembre, B., J. V. Arnekleiv & J. H. L'Abée-Lund, 2001. Effects of water discharge and temperature on
 474 the seaward migration of anadromous brown trout, *Salmo trutta*, smolts. *Ecology of Freshwater Fish*
 475 10: 61-64.

476 Hesthagen, T. & E. Garnås, 1986. Migration of Atlantic salmon smolts in River Orkla of central
 477 Norway in relation to management of a hydroelectric station. *North American Journal of Fisheries*
 478 *Management* 6: 376-382.

479 Holmes, R., J. W. Hayes, W. Jiang, A. Quarterman & L. N. Davey, 2014. Emigration and mortality of
 480 juvenile brown trout in a New Zealand headwater tributary. *Ecology of Freshwater Fish* 23: 631-643.

481 Huntingford, F. A., J. E. Thorpe, C. G. de Leaniz & D. W. Hay, 1992. Patterns of growth and smolting in
 482 autumn migrants from a Scottish population of Atlantic salmon, *Salmo salar* L. *Journal of Fish Biology*
 483 41:43-51.

484 Hvidsten, N. A. & L. P. Hansen, 1989. Increased recapture rate of adult Atlantic salmon, *Salmo salar*
 485 L., stocked as smolts at high water discharge. *Journal of Fish Biology* 32: 153-154.

486 Ibbotson, A. T., W. D. Riley, W. R. C. Beaumont, A. C. Cook, M. J. Ives, A. C. Pinder & L. J. Scott, 2013.
 487 The source of autumn and spring downstream migrating juvenile Atlantic salmon in a small lowland
 488 river. *Ecology of Freshwater Fish* 22: 73-81.

489 Jensen, A. J., B. Finstad, P. Fiske, N. A. Hvidsten, A. H. Rikardsen & L. Saksgård, 2012. Timing of smolt
 490 migration in sympatric populations of Atlantic salmon (*Salmo salar*), brown trout (*Salmo trutta*), and
 491 Arctic char (*Salvelinus alpinus*). Canadian Journal of Fisheries and Aquatic Sciences 69: 711-723.

492 Jonsson, B., 1985. Life history patterns of freshwater resident and sea-run migrant brown trout in
 493 Norway. Transactions of the American Fisheries Society 114: 182-194.

494 Jonsson, B. & N. Jonsson, 2009. Migratory timing, marine survival and growth of anadromous brown
 495 trout *Salmo trutta* in the River Imsa, Norway. Journal of Fish Biology 74: 621-638.

496 Jonsson, B. & N. Jonsson, 2011. Ecology of Atlantic salmon and Brown Trout: Habitat as a template
 497 for life histories. Fish and Fisheries Series 33, Springer, Dordrecht, the Netherlands.

498 Jonsson, B., & J. Ruud-Hansen, 1985. Water temperature as the primary influence on timing of
 499 seaward migrations of Atlantic salmon (*Salmo salar*) smolts. Canadian Journal of Fisheries and
 500 Aquatic Sciences 42: 593-595.

501 Jonsson, N., 1991. Influence of water flow, water temperature and light on fish migration in rivers.
 502 Nordic Journal of Freshwater Research 66: 20-35.

503 Jonsson, N. & B. Jonsson, 2002. Migration of anadromous brown trout in a Norwegian river.
 504 Freshwater Biology 47: 1391-1401.

505 Jonsson, N. & B. Jonsson, 2014. Time and size at seaward migration influence the sea survival of
 506 Atlantic salmon (*Salmo salar* L.). Journal of Fish Biology 84: 1457-1473.

507 Keeley, E. R., 2001. Demographic responses to food and space competition by juvenile steelhead
 508 trout. Ecology 82: 1247-1259.

509 Klemetsen, A., P. A. Amundsen, J. B. Dempson, B. Jonsson, N. Jonsson, M. F. O'Connell & E.
 510 Mortensen, 2003. Atlantic salmon *Salmo salar* L., brown trout *Salmo trutta* L. and Arctic charr
 511 *Salvelinus alpinus* (L.): a review of aspects of their life histories. Ecology of Freshwater Fish 12: 1-59.
 512 Larsen, M. H., A. N. Thorn, C. Skov & K. Aarestrup, 2013. Effects of passive integrated transponder
 513 tags on survival and growth of juvenile Atlantic salmon *Salmo salar*. Animal Biotelemetry 1: 1-7.
 514 Lucas, M. C. & E. Baras, 2001. Migration of Freshwater Fishes. Blackwell Science Ltd, Oxford.
 515 Marine Institute 2014. Newport Research Facility, Annual Report No. 58, 2013.
 516 McCormick, S. D., L. P. Hansen, T. P. Quinn & R. L. Saunders, 1998. Movement, migration, and
 517 smolting of Atlantic salmon (*Salmo salar*). Canadian Journal of Fisheries and Aquatic Sciences 55: 77-
 518 92.
 519 McGinnity, P., E. de Eyto, T. F. Cross, J. Coughlan, K. Whelan & A. Ferguson, 2007. Population specific
 520 smolt development, migration and maturity schedules in Atlantic salmon in a natural river
 521 environment. Aquaculture 273: 257-268.
 522 Nathan, R., W. M. Getz, E. Revilla, M. Holyoak, R. Kadmon, D. Saltz & P. E. Smouse, 2008. A
 523 movement ecology paradigm for unifying organismal movement research. PNAS 105: 19052-19059.
 524 Økland, F., B. Jonsson, A. J. Jensen & L. P. Hansen, 1993. Is there a threshold size regulating seaward
 525 migration of brown trout and Atlantic salmon? Journal of Fish Biology 42: 541-550.
 526 Pinder, A. C., W. D. Riley, A. T. Ibbotson & W. R. C. Beaumont, 2007. Evidence for an autumn
 527 downstream migration and the subsequent estuarine residence of 0+ year juvenile Atlantic salmon,
 528 *Salmo salar* L., in England. Journal of Fish Biology 71: 260-264.

529 Quéméré, E., C. Perrier, A.-L. Besnard, G. Evanno, J.-L. Baglinière, Y. Guiguen & S. Launey, 2014. An
 530 improved PCR-based method for faster sex determination in brown trout (*Salmo trutta*) and Atlantic
 531 salmon (*Salmo salar*). Conservation Genetics Resources, 6: 825-827.

532 R Core Team, 2014. R: A language and environment for statistical computing. R Foundation for
 533 Statistical Computing, Vienna, Austria, <http://www.R-project.org/>.

534 Richards, S. A., 2008. Dealing with overdispersed count data in applied ecology. Journal of Applied
 535 Ecology, 45: 218–227.

536 Riddell, B. E., & W. C. Leggett, 1981. Evidence of an adaptive basis for geographic variation in body
 537 morphology and time of downstream migration of juvenile Atlantic salmon (*Salmo salar*). Canadian
 538 Journal of Fisheries and Aquatic Sciences 38: 308-320.

539 Riley, W. D., A. T. Ibbotson, N. Lower, A. C. Cook, A. Moore, S. Mizuno, A. C. Pinder, W. R. C.
 540 Beaumont & L. Privitera, 2008. Physiological seawater adaptation in juvenile Atlantic salmon (*Salmo*
 541 *salar*) autumn migrants. Freshwater Biology 53: 745-755.

542 Riley, W. D., A. T. Ibbotson & W. R. C. Beaumont, 2009. Adult returns from Atlantic salmon, *Salmo*
 543 *salar*, parr autumn migrants. Fisheries Management and Ecology 16: 75-76.

544 Taal, I., M. Kesler, L. Saks, M. Rohtla, A. Verliin, R. Svirgsden, K. Jurgens, M. Vetemaa & T. Saat, 2014.
 545 Evidence for an autumn downstream migration of Atlantic salmon *Salmo salar* (Linnaeus) and brown
 546 trout *Salmo trutta* (Linnaeus) parr to the Baltic Sea. Helgoland Marine Research 68: 373-377.

547 Tanguy, J., D. Ombredane, J. Bagliniere & P. Prunet, 1994. Aspects of parr-smolt transformation in
 548 anadromous and resident forms of brown trout (*Salmo trutta*) in comparison with Atlantic salmon
 549 (*Salmo salar*). Aquaculture 121: 51-63.

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550 Taylor, L. R. & R. A. J. Taylor, 1977. Aggregation, migration and population mechanics. Nature 265:
551 415-421.

552 Venables, W. N., & B. D. Ripley, 2002. Modern Applied Statistics with S. Fourth Edition. Springer,
553 New York. ISBN 0-387-95457-0

554 ~~Villar Guerra, D. del, K. Aarestrup, C. Skov & A. Koed, 2014. Marine migrations in anadromous brown~~
555 ~~trout (*Salmo trutta*). Fjord residency as a possible alternative in the continuum of migration to the~~
556 ~~open sea. Ecology of Freshwater Fish 23: 594-603.~~

557 Wolf, P. A., 1951. A trap for the capture of fish and other organisms moving downstream.
558 Transactions of the American Fisheries Society 80: 41-45.

559 Wysujack, K., L. A. Greenberg, E. Bergman & I. C. Olsson, 2008. The role of the environment in partial
560 migration: food availability affects the adoption of a migratory tactic in brown trout *Salmo trutta*.
561 Ecology of Freshwater Fish 18: 52-59.

562 Youngson, A. F., R. J. G. Buck, T. H. Simpson & D. W. Hay, 1983. The autumn and spring emigrations
563 of juvenile Atlantic salmon, *Salmo salar* L., from the Girnock Burn, Aberdeenshire, Scotland:
564 environmental release of migration. Journal of Fish Biology 23: 625-639.

565 Zydlewski, G. B., A. Haro & S. D. McCormick, 2005. Evidence for cumulative temperature as an
566 initiating and terminating factor in downstream migratory behaviour of Atlantic salmon (*Salmo*
567 *salar*) smolts. Canadian Journal of Fisheries and Aquatic Sciences 62: 68-78.

568 Zydlewski, G. B., G. Horton, T. Dubreuil, B. Lecther, S. Casey & J. Zydlewski, 2006. Remote
569 monitoring of fish in small streams: a unified approach using PIT tags. Fisheries 31: 492-502.

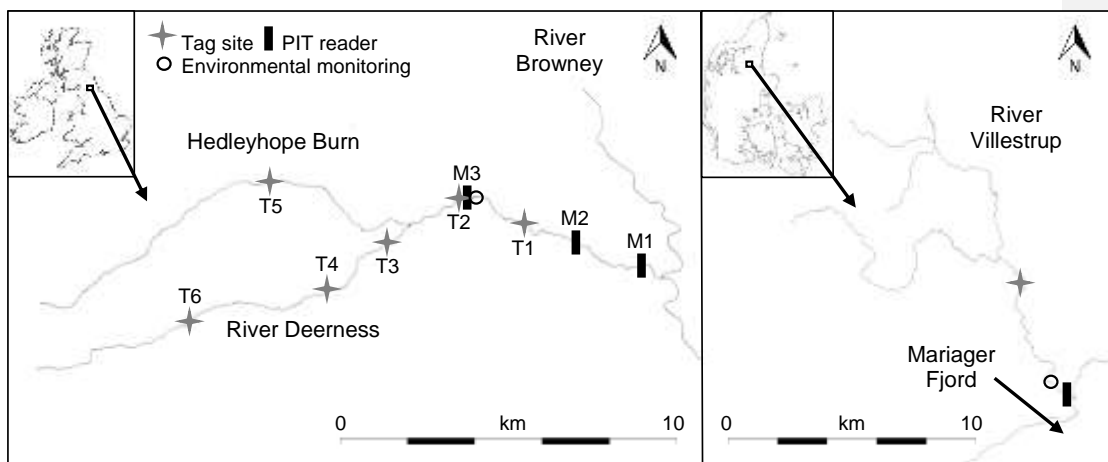
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Fig. 1 Map of the Deerness and Villestrup study areas, showing the locations of tagging sites, fixed PIT readers and environmental monitoring stations with stars, thick lines and open circles, respectively. On the Deerness, an environmental monitoring station was also placed at M3. Inset maps show the location of the study areas nationally.

Fig. 2 Daily detection frequency of PIT tagged trout parr detected downstream of the site at which they were tagged and released on the Deerness (a) and Villestrup (c). Date of first detection only is displayed. Shaded regions represent periods of minimal or no detection efficiency due to PIT antennae malfunction. Individuals detected prior to 1 Feb were labelled autumn migrants and those detected from 1 Feb labelled spring smolts following published convention (Ibbotson et al. 2013). In addition, a downstream trap was operational from 18 March on the Villestrup, indicated by the arrow on panel (c). Average daily water temperature (solid lines) and average daily water level (dotted lines) are also displayed for the Deerness (b) and Villestrup (d)

Fig. 3 The mean length, mass and condition factor at tagging (\pm SEM) of undetected trout, autumn downstream migrants, spring downstream migrants and upstream migrants in the Deerness (top row) and the Villestrup (bottom row)

Fig. 4 The mean net ground speed (\pm SEM) of autumn and spring migrants in the Deerness.



[Fig. 1](#)

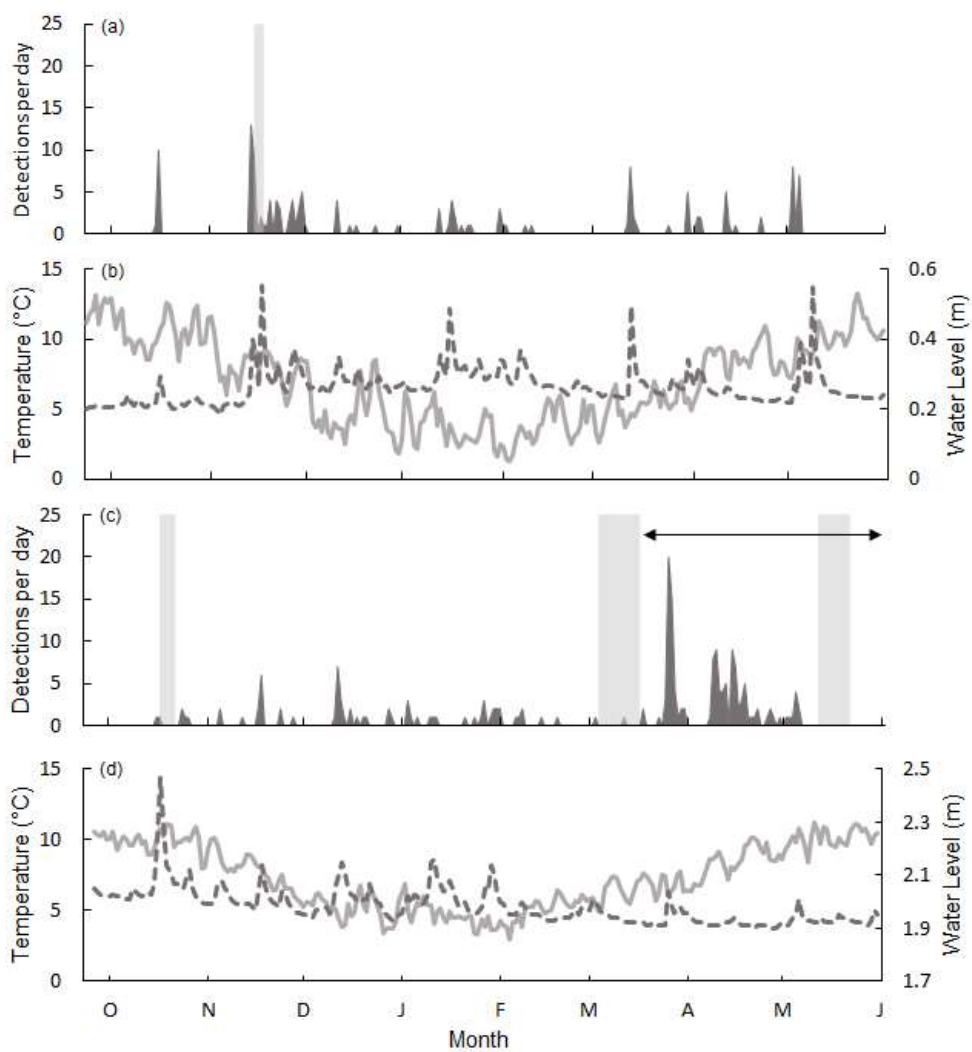


Fig. 2

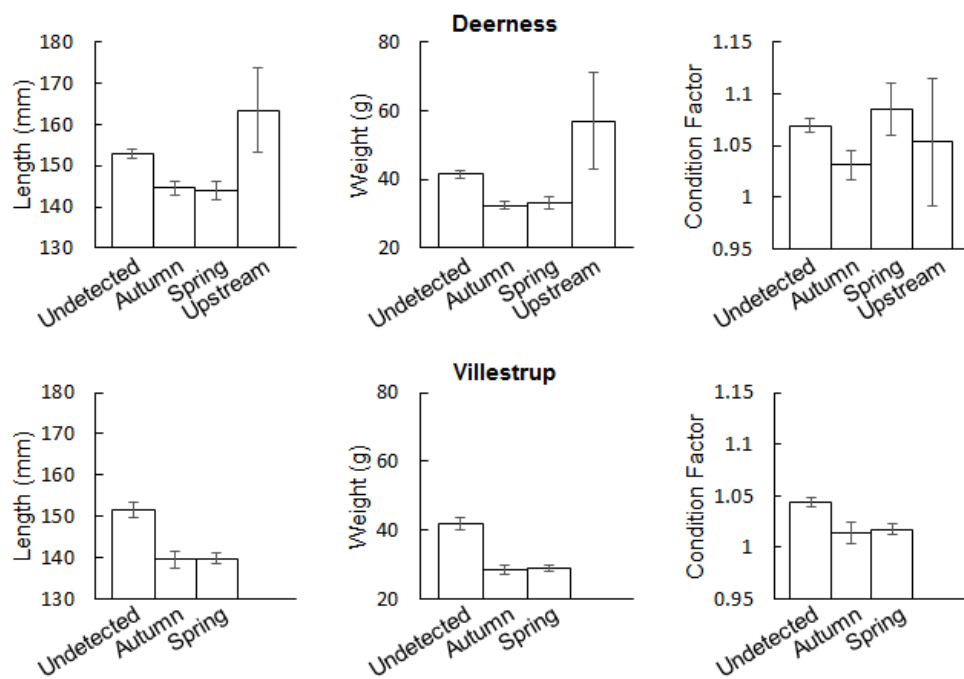


Fig. 3

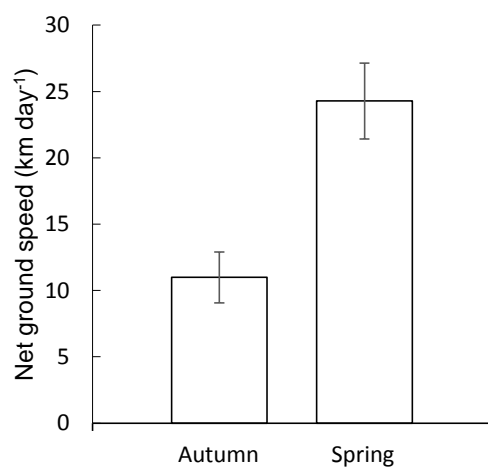


Fig. 4

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Table 1: Details of tag site-specific variables on the Deerness, along with results of PIT telemetry and March recapture surveys by site.

<u>Tag Site</u>	<u>Distance from river mouth (km)</u>	<u>Density '14 (n·100m⁻²)</u>	<u>Density '15 (n·100m⁻²)</u>	<u>%Δ Density</u>	<u>No. fish tagged</u>	<u>% Migrated Downstream</u>	<u>% Recaptured residents</u>	<u>% Loss from mortality or local dispersal</u>
<u>T1</u>	<u>4.2</u>	<u>12.0</u>	<u>14.3</u>	<u>+19.2</u>	<u>214</u>	<u>33.6</u>	<u>4.2</u>	<u>62.2</u>
<u>T2</u>	<u>6.2</u>	<u>9.2</u>	<u>3.7</u>	<u>-59.8</u>	<u>99</u>	<u>10.1</u>	<u>6.1</u>	<u>83.8</u>
<u>T3</u>	<u>8.6</u>	<u>12.2</u>	<u>1.9</u>	<u>-84.4</u>	<u>91</u>	<u>28.6</u>	<u>3.3</u>	<u>68.1</u>
<u>T4</u>	<u>10.8</u>	<u>9.5</u>	<u>1.5</u>	<u>-84.2</u>	<u>55</u>	<u>18.2</u>	<u>7.3</u>	<u>74.5</u>
<u>T5</u>	<u>11.8</u>	<u>12.2</u>	<u>2.3</u>	<u>-81.1</u>	<u>140</u>	<u>12.1</u>	<u>2.1</u>	<u>85.8</u>
<u>T6</u>	<u>14.5</u>	<u>7.7</u>	<u>7.5</u>	<u>-2.6</u>	<u>67</u>	<u>7.5</u>	<u>10.4</u>	<u>82.1</u>

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Table 2: Generalised linear model outputs of migration phenology analyses. All retained models within 6 ΔAIC are displayed. Significant variables are in bold, with values for predictor variables representing coefficient estimates.

Model	AICc	ΔAICc	df	Intercept	Water Level	Δ Water Level	Water Temperature	Photoperiod
<u>Deerness Autumn</u>								
<u>1</u>	<u>202.3</u>	<u>0.0</u>	<u>4</u>	<u>-12.00</u>	<u>33.30</u>		<u>0.24</u>	
<u>Deerness Spring</u>								
<u>1</u>	<u>166.8</u>	<u>0.0</u>	<u>3</u>	<u>-1.76</u>		<u>22.83</u>		
<u>2</u>	<u>168.3</u>	<u>1.5</u>	<u>4</u>	<u>-9.51</u>	<u>20.24</u>			<u>0.004</u>
<u>3</u>	<u>169.1</u>	<u>2.3</u>	<u>3</u>	<u>-5.84</u>	<u>17.35</u>			
<u>Villestrup Autumn</u>								
<u>1</u>	<u>207.7</u>	<u>0.0</u>	<u>4</u>	<u>-20.80</u>	<u>10.83</u>		<u>-0.34</u>	
<u>2</u>	<u>208.6</u>	<u>0.9</u>	<u>4</u>	<u>-18.23</u>	<u>11.02</u>			<u>-0.011</u>
<u>Villestrup Spring</u>								
<u>1</u>	<u>311.7</u>	<u>0.0</u>	<u>4</u>	<u>-1.87</u>		<u>27.04</u>	<u>0.20</u>	
<u>2</u>	<u>312.0</u>	<u>0.3</u>	<u>3</u>	<u>-0.26</u>		<u>21.68</u>		
<u>3</u>	<u>316.1</u>	<u>4.4</u>	<u>4</u>	<u>-29.64</u>	<u>13.77</u>			<u>0.004</u>
<u>4</u>	<u>316.4</u>	<u>4.7</u>	<u>4</u>	<u>-34.35</u>	<u>16.62</u>		<u>0.28</u>	
<u>5</u>	<u>317.3</u>	<u>5.6</u>	<u>3</u>	<u>-17.57</u>	<u>9.10</u>			

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Table 3: Generalised linear model outputs of migration tendency analyses. All retained models within 6 Δ AIC are displayed. Significant variables are in bold, with values for predictor variables representing coefficient estimates.

Model	AICc	Δ AICc	df	Intercept	Length	Weight	Condition	Density	Distance	Days
<u>Deerness Autumn</u>										
1	487.4	0.0	3	0.26		-0.032			-0.136	
2	489.7	2.3	5	4.29	-0.023		-1.86		-0.146	0.006
3	489.7	2.3	4	3.93	-0.022		-1.42		-0.137	
4	491.3	3.9	3	2.48	-0.022				-0.143	
<u>Deerness Spring</u>										
1	340.7	0.0	4	-1.35		-0.041	1.79		-0.178	
2	341.0	0.3	3	3.42	-0.029				-0.178	
3	343.0	2.3	3	0.24		-0.035			-0.164	
<u>Villestrup Autumn</u>										
1	353.9	0.0	3	2.15		-0.023	-3.33			
2	354.6	0.7	2	-1.21		-0.024				
3	356.0	2.1	3	4.34	-0.014		-4.19			
4	358.4	4.5	2	-0.002	-0.014					
5	359.3	5.4	2	2.34			-4.22			
<u>Villestrup Spring</u>										
1	527.0	0.0	3	4.70		-0.026	-4.42			
2	532.6	5.6	3	7.40	-0.017		-5.47			

612 Online Resource 1: Investigating the phenology of downstream migration in juvenile trout.

613 Methodology of Deerness recapture surveys in summer 2014, used to investigate summer-time

614 dispersal of trout parr.

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615 Following the initial PIT-tagging of trout parr at sites along the Deerness, recapture sessions

616 during the summer of 2014 enabled the re-sampling of individuals to determine their level of

617 movement. Three single-pass electrofishing surveys were conducted at each site in contiguous stop-

618 netted 20 m stream sections over a period of *ca.* two months (early July to mid September 2014).

619 The distance surveyed was increased in each session from initially 100 m to finally 200 m upstream

620 and downstream of each starting point. During the first recapture session at sites T2, T3 and T4, and

621 during the first two recapture sessions at sites T1, T5 and T6, any untagged individuals captured

622 were tagged and subsequently released back into the population. The frequencies of maximum

623 longitudinal distances covered by each recaptured fish from a previous site of release showed a

624 typical inverse-power distribution. Following the methods of Bubb et al. (2004), linear

625 transformation using a double-In plot enabled regression analysis. Upstream and downstream

626 dispersal were analysed separately and the regression lines compared. To maximise sample size, the

627 recorded dispersal ranges were combined from all study sites.

628 References

629 Bubb, D. H., T. J. Thom & M. C. Lucas, 2004. Movement and dispersal of the invasive signal crayfish

630 *Pacifastacus leniusculus* in upland rivers. Freshwater Biology, 49: 357-368.

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Online Resource 2: Investigating the phenology of downstream migration in juvenile trout.

Results and analysis of the Deerness recapture surveys of trout parr dispersal in summer 2014.

A total number of 330 fish were recaptured during single pass recapture surveys on the Deerness between late July and mid-September 2014. The maximum distance covered by any one trout tagged and recaptured within the same Deerness study site, July to September 2014, was 260 m, however many recaptured individuals (50.2%) remained within the 20m zone in which they were first released, and the majority (83.0%) travelled no further than 60m from previous known locations (Fig. S1). The probability, M , of moving a dispersal distance of D m was described by an inverse-power function using the inverse cumulative proportion of dispersers in each 20 m sample zone from the site of release (Bubb et al. 2004):

$$M = CD^{-n}$$

C and n are scaling constants. A highly significant negative relationship between $\ln M$, both upstream and downstream, and $\ln D$ was found (Upstream: $F_{1,7} = 82.7$, $p < 0.001$, $R^2 = 0.92$; Downstream: $F_{1,10} = 121.7$, $p < 0.001$, $R^2 = 0.92$; Fig. S2) under the equations:

$$\ln M (\text{upstream}) = 7.25 - 2.26(\ln D)$$

$$\ln M (\text{downstream}) = 7.22 - 2.21(\ln D)$$

No significant difference between the gradients of the two regression lines was found ($t_{20} = -0.18$, $p = 0.86$), meaning trout were equally likely to disperse upstream and downstream (Fig. S2).

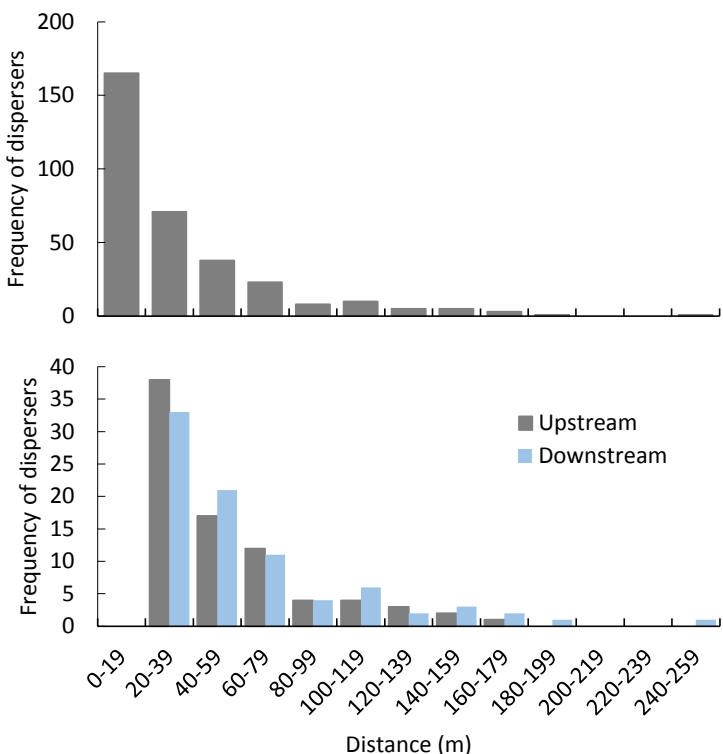


Fig. S1: Frequency distribution of the maximum longitudinal movements made by trout from a previous release site (top), with upstream and downstream movements separated (bottom), all study sites combined.

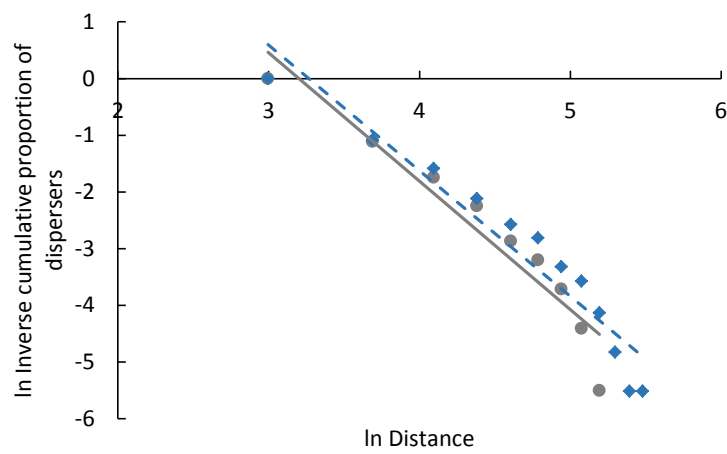


Fig. S2: Double-In plot of the inverse cumulative proportion of dispersers in upstream (grey) and downstream (blue) directions.

References

Bubb, D. H., T. J. Thom & M. C. Lucas, 2004. Movement and dispersal of the invasive signal crayfish *Pacifastacus leniusculus* in upland rivers. *Freshwater Biology*, 49: 357-368.

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